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Review

Indirect calorimetry in nutritional therapy. A position paper by the ICALIC study group

Taku Oshima ^a, Mette M. Berger ^b, Elisabeth De Waele ^c, Anne Berit Guttormsen ^{d, e, f},
 Claudia-Paula Heidegger ^g, Michael Hiesmayr ^h, Pierre Singer ⁱ, Jan Wernerman ^j,
 Claude Pichard ^{k, *}

^a Department of Emergency and Critical Care Medicine, Chiba University Graduate School of Medicine, 1-8-1 Inohana Chuou-ku, Chiba City, Chiba 260-8677, Japan

^b Adult Intensive Care, Lausanne University Hospital, 1011 Lausanne, Switzerland

^c Department of Intensive Care, Vrije Universiteit Brussel, Brussels, Belgium

^d Department of Anaesthesiology and Intensive Care, Haukeland University Hospital, Jonas Liesvei 65, 5021 Bergen, Norway

^e Department of Clinical Medicine University of Bergen, Bergen, Norway

^f Haukeland Universitetssykehus Laboratoriebygget, 7. etg. Heis øst, Norway

^g Service of Intensive Care, Geneva University Hospital, Rue Gabrielle-Perret-Gentil 4, 1211 Geneva 14, Switzerland

^h Division of Cardiac Thoracic Vascular Anesthesia and Intensive Care Medicine, University Hospital of Vienna, Waehringer Guertel 18-20, 1090 Vienna, Austria

ⁱ Critical Care Medicine, Institute for Nutrition Research, Rabin Medical Center, Beilison Hospital, Petah Tikva 49100, Israel

^j Department of Anesthesiology and Intensive Care Medicine, Karolinska University Hospital Huddinge, Sweden

^k Nutrition Unit, Geneva University Hospital, Rue Gabrielle-Perret-Gentil 4, 1211 Geneva 14, Switzerland

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SUMMARY

Background & aims: This review aims to clarify the use of indirect calorimetry (IC) in nutritional therapy for critically ill and other patient populations. It features a comprehensive overview of the technical concepts, the practical application and current developments of IC.

Methods: Pubmed-referenced publications were analyzed to generate an overview about the basic knowledge of IC, to describe advantages and disadvantages of the current technology, to clarify technical issues and provide pragmatic solutions for clinical practice and metabolic research. The International Multicentric Study Group for Indirect Calorimetry (ICALIC) has generated this position paper.

Results: IC can be performed in in- and out-patients, including those in the intensive care unit, to measure energy expenditure (EE). Optimal nutritional therapy, defined as energy prescription based on measured EE by IC has been associated with better clinical outcome. Equations based on simple anthropometric measurements to predict EE are inaccurate when applied to individual patients. An ongoing international academic initiative to develop a new indirect calorimeter aims at providing innovative and affordable technical solutions for many of the current limitations of IC.

Conclusion: Indirect calorimetry is a tool of paramount importance, necessary to optimize the nutrition therapy of patients with various pathologies and conditions. Recent technical developments allow broader use of IC for in- and out-patients.

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1. Introduction

Indirect calorimetry (IC) measures the oxygen consumption and the carbon dioxide production, which correspond to the cellular respiration and allows to calculate the energy expenditure (EE) of the whole body [1]. The study of the basic principles started more than 100 years ago mainly by physicists and chemists, from the discovery of gas and its components to the establishment of the

* Corresponding author. Geneva University Hospital, 1211 Geneva 14, Switzerland. Tel.: +41 22 372 93 45; fax: +41 22 372 93 63.

E-mail addresses: t.oshima@chiba-u.jp (T. Oshima), Mette.Berger@chuv.ch (M.M. Berger), Elisabeth.DeWaele@uzbrussel.be (E. De Waele), anne.guttormsen@helse-bergen.no (A.B. Guttormsen), claudia-paula.heidegger@hcuge.ch (C.-P. Heidegger), michael.hiesmayr@meduniwien.ac.at (M. Hiesmayr), psinger@clalit.org.il (P. Singer), jan.wernerman@karolinska.se (J. Wernerman), claudie.pichard@unige.ch (C. Pichard).

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concept of gas exchange related to combustion [1]. Technical progresses have enabled measurements of oxygen (O₂) and carbon dioxide (CO₂) concentrations, as well as the volume of respiratory gasses and heat production of living organisms. In 1949, Weir derived an equation to calculate EE based on the heat produced when a given substrate was burned and the volume of O₂ needed to burn this substrate and on the protein oxidation derived from urea excretion via the kidney. The EE can be derived from O₂ consumption, CO₂ production and urea excretion [Table 1] [2]. Many proposed formulae deviate minimally from the Weir formula, and are not valid if other substrates such as ketones or pyruvate are oxidized in substantial amounts [5]. The principle that nitrogen is neither utilized nor produced during respiration has enabled calculation of EE without measuring inhaled air volume [4]. This principle, known as the Haldane transformation, has contributed greatly to simplifying the measurement systems as only expiratory gas volume needed to be measured.

Measuring EE was labor-intensive and reserved for laboratory research. It is only in the 1980's, that indirect calorimeters were commercialized for medical use. Their complexity and high cost have limited their use for clinical routine during the last 4 decades. IC was mostly used for metabolic research. Nevertheless, in clinical practice it has been taking hold, and became indispensable in pediatric intensive care units (ICU) [6]. In critically ill adults, half of patients in a mixed medical-surgical ICU had indications for IC [7,8], but were rarely measured.

The assessment of EE requires IC and cannot be predicted by equations [9,10]: a number of predictive equations based on simple anthropometric measures and some including gender, age and minute ventilation have been proposed for clinical use as a surrogate of measuring EE [11]. Unfortunately, many reports have shown that predictive equations are not accurate enough, because patients with acute or chronic conditions have different metabolic characteristics, reflected by highly variable EE. Body composition is also an important modifier of EE because fat free mass accounts for most of EE [12].

Recently, it became clear that both overfeeding and underfeeding can be harmful [13–18] and that optimizing nutrition support to the patients specific needs is an urgent task. IC is the only practical clinical method to measure EE of in- and out-patients [3,19,20] in order to tailor nutrition therapy/support to their specific needs; assuming that the energy target should match EE. However, the calorimeters currently available on the market are not sufficiently accurate [9,21–24], difficult to use, and too expensive to be readily accessible in general hospitals.

Table 1
Equations used for the calculations related to indirect calorimetry [2–4].

<i>Calculations of O₂ consumption and CO₂ production</i>
$VO_2 = (Vi \times FiO_2) - (Ve \times FeO_2)$
$VCO_2 = (Ve \times FeCO_2) - (Vi \times FiCO_2)$
<i>Haldane transformation</i>
Assumption based on the concept that N ₂ is constant in inspired and expired gas
$Vi = [FeN_2/FiN_2] \times Ve$
$FeN_2 = (1 - FeO_2 - FeCO_2)$
$FiN_2 = (1 - FiO_2 - FiCO_2)$
If FiCO ₂ of 0.03–0.05% is ignored,
$VO_2 = [(1 - FeO_2 - FeCO_2) \times (FiO_2 - FeO_2) \times Ve] / (1 - FiO_2)$
<i>Weir's equation</i>
$EE = [(VO_2 \times 3.941) + (VCO_2 \times 1.11) + (uN_2 \times 2.17)] \times 1.44$

VO₂: O₂ consumption (L/min), VCO₂: CO₂ production (L/min), Vi: inspired volume (L), Ve: expired volume (L), FiO₂: fraction of inspired oxygen, FeO₂: fraction of expired oxygen, FeN₂: fraction of expired nitrogen, FiN₂: fraction of inspired nitrogen, EE: energy expenditure (kcal/d), uN₂: urinary nitrogen (g/d).

An ongoing initiative supported by two major academic organizations (i.e. The European Society for Clinical Nutrition and Metabolism (ESPEN) and The European Society for Intensive Care Medicine (ESICM)) was launched to develop a new indirect calorimeter. The goals were defined by a bottom-up approach with the aim of developing an accurate, easy-to-use and affordable indirect calorimeter for the use of the scientific and medical community.

This review aims at summarizing the scientific background supporting IC in order to optimize nutrition therapy for critically ill patients and other patient populations. It features a comprehensive overview of the technical concepts, the practical application and current developments of IC.

2. Technical concepts

2.1. Calorimetry: the basics

IC measures inspired and expired gas exchanges to calculate EE. This is possible because heat production is tightly correlated with O₂ consumption (VO₂) and CO₂ production (VCO₂) according to the type of energy substrate [20,25].

The conditions of the subjects during IC must be defined as they deeply influence the results. For healthy individuals, basal energy expenditure (BEE) is measured in a resting state that is free of physical and psychological stress, a thermally neutral environment, i.e. at temperature ranges where energy used for the body temperature maintenance is minimal, and a fasting state, i.e. no oral intake for more than 10 h prior to the measurement, to avoid the EE related to physical activity and diet-induced thermogenesis (DIT) [Table 2]. DIT is defined as the production of heat related to substrate oxidation during energy uptake. Resting energy expenditure (REE) is defined as the sum of BEE and DIT, and total energy expenditure (TEE) as the sum of REE and activity induced energy expenditure (AEE) [1,25]. By definition, BEE measurements must be conducted in conditions that are unfeasible for diseased individuals [Table 3]. In clinical practice, REE or TEE reflects the patient energy needs. For patients in the ICU, measured EE should be considered as TEE. If physical activity becomes a standard routine ICU care in the future, then this statement must be revised.

2.2. How is EE measured?

IC requires the measurement of inspired and expired O₂ and expired CO₂ concentrations, as well as the volume of expired gas per minute to calculate the VO₂(L/min) and VCO₂(L/min) [28]. Then VO₂ and VCO₂ are used to calculate the EE(kcal/day) using the Weir's equation [Table 1] [2,20,28,29].

In a mechanically ventilated patient, the gas sampling is obtained from the circuit connecting the endotracheal tube to the ventilator, and measured by using either the breath-by-breath analysis [Fig. 1 a] or the analysis using a mixing chamber [Fig. 1 b] [Table 4]. In spontaneously breathing subjects, a ventilated canopy hood or a fitted face mask is used to collect the inspired and expired gas [Fig. 1c] [28]. Air leaks of respiratory gases alter the accuracy of the measurements and should be avoided.

For measurements using the canopy without O₂ enrichment, VO₂ and VCO₂ can be calculated as a difference between the O₂ concentration in ambient air and the measured O₂ and CO₂ concentration in the expired gas, collected by the canopy. For measurements in mechanically ventilated conditions or using the canopy with O₂ enrichment, the measurements are more complex. Breath-by-breath systems measure the exhaled gas volume and the O₂ and CO₂ concentration transitions, and integrate the product of instantaneous expired gas concentrations with instantaneous expiratory flow over time. A mixing chamber system measures the

Table 2
Components of the energy expenditure in healthy subjects and diseased individuals [1].

Components of energy expenditure	Definition
Basal energy expenditure (BEE)	Energy expended in fasting state, resting in lying position at neutral ambient temperature, free of physical and psychological stress. Note: <u>Only applicable in healthy subjects.</u>
Diet-induced thermogenesis (DIT)	Oxidation of energy substrates during oral, enteral or intravenous energy intake
Activity energy expenditure (AEE)	Energy expenditure to support physical activity
Resting energy expenditure (REE)	BEE + DIT
Total energy expenditure (TEE)	REE + AEE

inhaled and exhaled gasses separately, to detect the global change in the inhaled and exhaled gas [30]. The expired volume is usually measured by a separate flow meter, or by a dilution technique using a constant flow chamber to calculate the volume [30]. Both systems use the Haldane transformation, i.e. the method to calculate inspired gas volume by the ratio of the inspired and expired nitrogen concentrations, to calculate the inspired gas volume thus simplifying the flow or volume measurements [Table 1] [4,20,28].

Some commercially available simplified devices only measures either VO_2 or the VCO_2 to calculate EE by assuming that the RQ is a fixed value (i.e. 0.8–0.85) [28,31,32]. While this type of assumption may be acceptable in healthy subjects on balanced nutrition [28], it is not recommended for patients because their substrate oxidation may change significantly according to the type of disease and nutrition. Assuming a fixed RQ in patients give inaccurate EE, in turn leading to suboptimal energy prescription. Using the VCO_2 and RQ of prescribed nutrition formulas (food quotient) to calculate EE has been proposed as a way to improve the accuracy of the calculation for ICU patients [32]. The analysis was conducted on stabilized patients who tolerated more than two-thirds of the prescribed nutrition allowing the mean EE bias of 7.7% ($=+141$ kcal/d) while improving the precision compared to predictive equations. However, the accuracy level of this method for individual patients can only be validated by conducting IC. Thus, this method can be considered as an alternative for predictive equations, but should not be considered as a valid alternative for IC in the general ICU population.

2.3. The reference device of the 20th century

Numerous indirect calorimeters have been in and out of the market in the past decades. However, the Deltatrac Metabolic

Monitor[®] (Datex, Finland) produced 35 years ago is often viewed as the reference device [9,22–24,33]. This device features both canopy and ventilator measurements [30]. When on ventilator mode, it uses the mixing chamber technique with a unique constant flow chamber to dilute the exhaled gas to enable calculations of VO_2 , VCO_2 and EE without directly measuring the expired gas volume [30]. The device has been repeatedly validated, including a comparison against mass spectrometry [30,34,35]. However, existing units are progressively disappearing and the manufacturer no longer offers any support.

2.4. Technology of modern indirect calorimeters

Calorimeters are designed to measure spontaneously breathing patients or mechanically ventilated patients [28]. The different techniques predetermine the limitations of their performances [Table 4].

Devices with breath-by-breath technology can be made smaller as they do not require a bulky mixing chamber. They generate rapid readings by measuring short intervals of gas samples, a valuable feature in case of exercise physiology or rapid shift in substrates oxidation.

Devices with a mixing chamber generate more stable measurements because the gases are physically “averaged” before being analyzed, allowing the gas analyzers to generate very accurate analysis. The mixing chamber typically occupies 3–5 L of space, precluding the making of a small device. The capacity to make reliable measurements in a short duration (e.g. 3–5 min) is also limited, as it takes just as much time for the gas concentrations in the mixing chamber to stabilize.

2.5. Accuracy and reproducibility

Three components of the hardware play a major role: the O_2 and CO_2 analyzers, and the flowmeter. Their accuracy, precision and reproducibility are critical for IC and are influenced by many factors [Table 5]. For breath-by-breath systems, the reaction time of the gas analyzers is important. The reliability of the software to synchronize the signals from the gas analyzers and the expiratory flowmeter to allow continuous calculations is a challenging demand. Small errors in the alignment of the acquired data can lead to great differences in the results. Mixing chamber devices are not as technically demanding. However, the use of the Haldane transformation formula introduces a mathematical limitation, especially in case of O_2 enrichment higher than 60% as the inaccuracy of the analyzers will be enhanced by the calculation [28,30,34].

Outside the calorimeter itself, the collection of inspired and expired gases by an appropriate and airtight system is mandatory [Fig. 2]. Avoiding leaks of inspired and expired gas is crucial, and

Table 3
Required conditions for accurate measurement of energy expenditure in healthy subjects or diseased individuals [1,26,27].

Parameter	Condition	Subject
BEE	At least 10 h after the previous meal Free of drugs Resting in supine position and free of physical stress Awake and free of psychological stress Normal body temperature Ambient temperature in zone of neutrality (27–29 °C)	Only healthy subjects
REE	At least 5 h after the previous meal, or under continuous feeding Minimum 2 h after alcohol and nicotine ingestion, 4 h after caffeine ingestion After 30 min of resting period Resting in supine position and free of physical stress Awake and free of psychological stress Comfortable environmental condition	Healthy subjects or patients
TEE	No specific conditions	Healthy subjects or patients

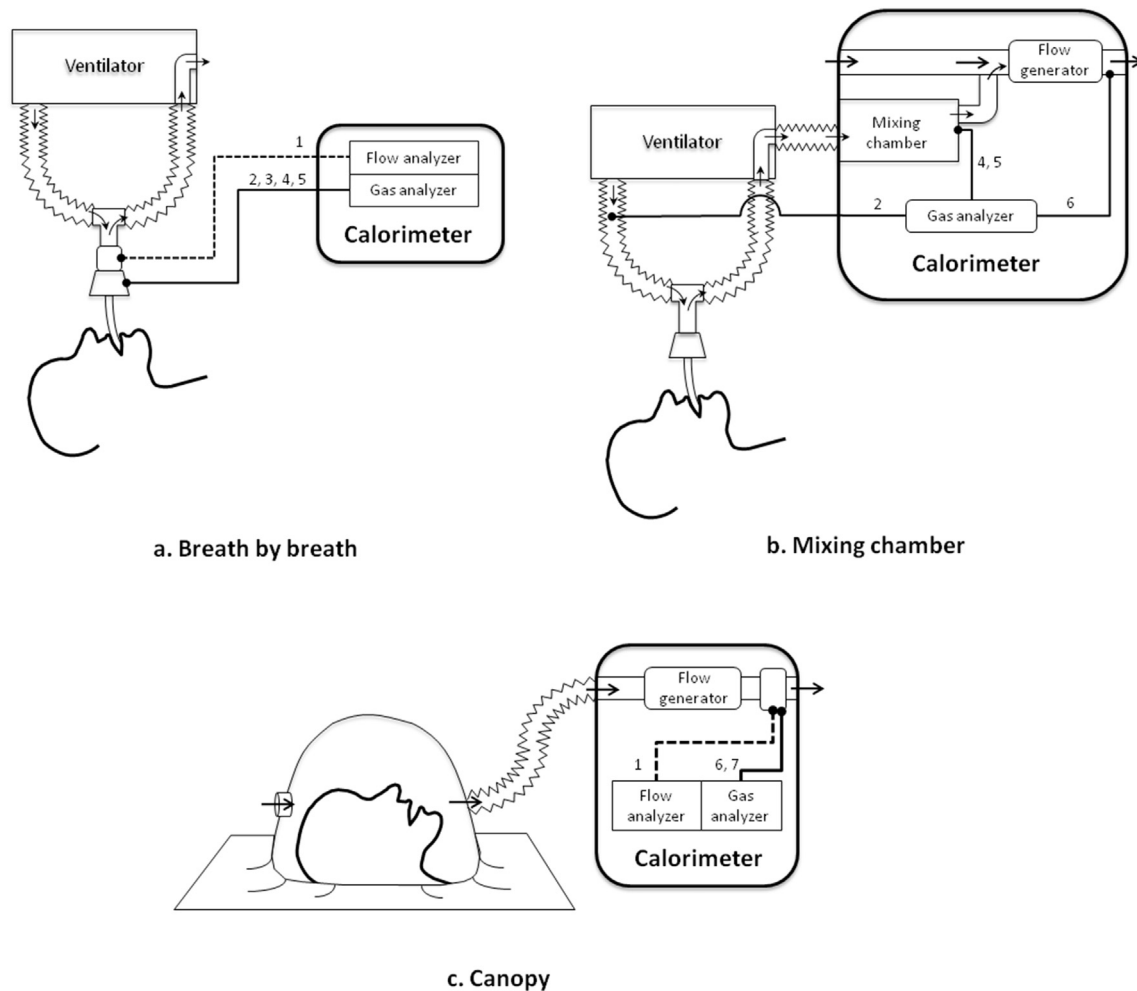


Fig. 1. Schematic presentation of indirect calorimetry used in patients on mechanical ventilation and on those breathing spontaneously. a) Breath by breath: Respiratory gas composition and flow are measured continuously by connecting the gas analyzers to the ventilator circuit. The signals received by the gas analyzers and flow meters are synchronized to calculate the oxygen consumption (VO_2 L/min) and CO_2 production (VCO_2 L/min) as the difference between the volumes of inhaled and exhaled O_2 and CO_2 per breath by integral calculations. The Haldane transformation [Table 1] is used to calculate the inhaled gas volume from exhaled gas volume measurement. The Weir's equation [Table 1] is used to calculate EE (kcal/d) per breath, and averaged for the duration of the measurement. The system is highly responsive to the dynamic changes of the EE, but prone to errors due to the response time of the gas analyzers and software. b) Mixing chamber: The O_2 concentration of inhaled air (FiO_2) is first measured. Exhaled gas is collected into the mixing chamber, where it is physically averaged and analyzed for O_2 (FeO_2) and CO_2 (FeCO_2) concentrations. The collected gas is eliminated through an independent chamber where the gas flow (Q) is kept constant at 40–45 L/min, to dilute the exhaled gas from the mixing chamber with the ambient air. CO_2 in the diluted gas (FedCO_2) is measured to calculate the CO_2 production (VCO_2 , L/min) by multiplying the concentration by the flow ($\text{VCO}_2 = \text{FedCO}_2 \times Q$). An equation using the Haldane transformation allows the calculation of the respiratory quotient (RQ) from the measured O_2 and CO_2 values ($\text{RQ} = (1 - \text{FiO}_2) / ((\text{FiO}_2 - \text{FeO}_2) / \text{FeCO}_2 - \text{FiO}_2)$), and thus enables the calculation of the oxygen consumption (VO_2 , L/min; $\text{VO}_2 = \text{VCO}_2 / \text{RQ}$). This unique method used in the Deltatrac Metabolic Monitor® (Datex, Finland) enables VO_2 and VCO_2 measurements without measuring the flow of the exhaled gas, which usually introduces technical difficulties. c) Canopy: The canopy is used to measure EE in spontaneously breathing subjects. The subject is placed under a clear canopy with a plastic drape to avoid air leakage. Calorimeters feature constant flow generator to create an outward flow through the canopy. The exhaled breath by the subject is diluted by the constant flow Q (L/min), and collected by the calorimeter for gas analysis (FedO_2 , FedCO_2), and enables calculations of VO_2 and VCO_2 [Table 1]. FiO_2 and FiCO_2 are either assumed as ambient air values or measured, depending on the calorimeter. These values are used to calculate the EE using the Weir's equation. (1: flow analysis, 2: FiO_2 , 3: FiCO_2 , 4: FeO_2 , 5: FeCO_2 , 6: FedCO_2 , 7: FedO_2 ; small arrows: respiratory gas flow, solid line: gas sampling, dotted line: signal for flow analysis, small arrows: respiratory gas flow, bold arrow: constant flow).

Table 4
Technologies used in commercially available calorimeters.

Patient condition	Patient application	Technology	Practical characteristics (advantages/disadvantages)
Spontaneous Breathing	Canopy	Constant flow dilution	Patient discomfort minimum Difficult to measure with O_2 supplementation
	Facemask	Breath by breath or mixing chamber	Supports O_2 and mask ventilation Patient discomfort, risk of leak
Mechanical Ventilation	In-circuit	Breath by breath	Small device, fast response Prone to error in calculation, dead space & resistance of the measurement components
	Gas collection	Mixing chamber	Stable measurements, validated in literature Large devices, difficult to disinfect

becomes even more critical in case of O₂ enrichment. High quality of calibration gas for calibration and periodic maintenance of the calorimeters guarantee the technical performances [25].

2.6. Alternative methods to measure energy expenditure

EE can be measured by four different methods: 1) direct calorimetry, 2) indirect calorimetry, 3) thermodilution (Fick method), and 4) ²H/¹H and ¹⁸O/¹⁶O doubly labeled water.

2.6.1. Direct calorimetry

Direct calorimetry is the direct measurement of heat production in the body. This concept is based on the phenomenon that all energy substrates, upon oxidation, produce heat. The subject needs to be confined in an insulated chamber to measure the heat production. The subjects also have to be able to maintain a complete resting state during the measurement in order to avoid extra heat production by physical activity. Thus the conditions are unrealistic for clinical use, and the availability is limited to a few specialized research centers.

2.6.2. Fick method

The Fick method requires a pulmonary artery catheter to measure the cardiac output, using the thermodilution method. Arterial and mixed venous oxygen contents must also be measured. After measuring the O₂ content in arterial and mixed venous blood from the pulmonary artery, VO₂ can be calculated using the Fick equation [Table 6]. The EE is calculated by assuming a fixed RQ. Several problems limit its use in clinical practice. First, only few patients have pulmonary artery catheters and the insertion of the catheter only for EE measurements would be too invasive. Second, the VO₂ calculated by this method is only a snapshot of the moment of the measurement, while the error of the thermodilution method is about 15% due to cardiac output variation over the respiratory cycle. Furthermore, mixed venous oxygen concentration may be overestimated because of the shunting of arterial blood from bronchial vessels, thus leading to underestimation of VO₂ and subsequently the EE.

2.6.3. Doubly labeled water

Water containing non-radioactive isotope labeled hydrogen and oxygen atoms (²H/¹H and ¹⁸O/¹⁶O) is given orally, after a baseline evaluation of the body liquids; urine, saliva, and blood. The evaluation of the body liquids is repeated after 7–12 days to calculate the variations of concentrations of the isotopes over time. CO₂ production can be calculated by observing the elimination rates of the isotopes from the body liquids. EE can be calculated by assuming a given RQ. The calculations are based on several assumptions such as steady-state CO₂ and H₂O turnover, and constant body water pool size during the measurement period. These assumptions may not be applicable for critically ill patients, as fluid volume shifts together with large changes in CO₂ production are frequently observed [3]. The costs of the doubly labeled water and of mass spectrometry measurements are very high. This method allows to

calculate EE, but the delay to obtain the results limits its use to research [3].

In summary, these three methods are too invasive, cumbersome or costly. IC remains the most practical method that is applicable in patients with various characteristics.

3. Practical considerations

3.1. Indications and limitations

IC is a non-invasive technique [25] applicable to many patients in order to individualize their nutrition therapy or for research purpose. However, IC may require special considerations for the interpretation of the results in a number of specific situations [Table 7].

3.1.1. Patients

By definition, the most important condition is the absence of air leak in the respiratory circuit. For example, patients with air-leaking chest drainage cannot be studied by IC [27]. Mechanically ventilated patients with high pressure settings on the ventilator are prone to air leakage at the level of the endotracheal tube [36]. IC in patients with unstable conditions is less useful, as the measurement will not represent their true metabolic characteristics. For example, agitated patients or those with seizures or other involuntary movements are difficult to assess, as measurements will include the EE related to the body movements, by nature inconstant, and therefore will not represent the true daily EE [27]. A patient should be in a resting condition or at least be able to keep calm during most of the IC duration [26], for the results to be a true representative value of the resting EE. Patients with unstable body temperature, variable pH due to CO₂ accumulation or other causes are also likely to present unreliable results, and measurements should be repeated periodically or after stabilization.

3.1.2. Treatments

Mechanical ventilation with FiO₂ >60% is likely to generate inaccurate measurements because of the Haldane transformation [27,30]. Patients on organ support treatments that supply O₂ to the blood or remove CO₂ from the blood (e.g. ECMO) and treatments that alter acid-base homeostasis (e.g. renal replacement therapy and liver support therapies such as Molecular Adsorbent Recirculating System (MARS)) also need special consideration.

Currently available IC devices do not provide valid solutions for these special conditions. However, the improvements of organ support therapies have enabled their frequent application in longer durations for patients under extremely severe conditions or waiting organ transplantation. Thus, technical solutions to conduct IC accurately in these patients are mandatory. De Waele et al. suggested a method for conducting IC in ECMO patients [37]. Development of commercial calorimeters designed for use with various treatment conditions will contribute to the improved nutrition therapy in these patients.

Table 5

Source of errors for indirect calorimetry depending on the measurement technology.

Technology	Specific factors	Common factors
Canopy	Leak of the gas collection Reliability of the constant flow	Accuracy of O ₂ , CO ₂ , and flow analyzers
Breath-by-breath	Response speed of gas analyzers Accuracy of data synchronization by the software	Haldane transformation introduces high variability when FiO ₂ >60%
Mixing chamber	Leak of the gas collection	Adequate maintenance and calibration

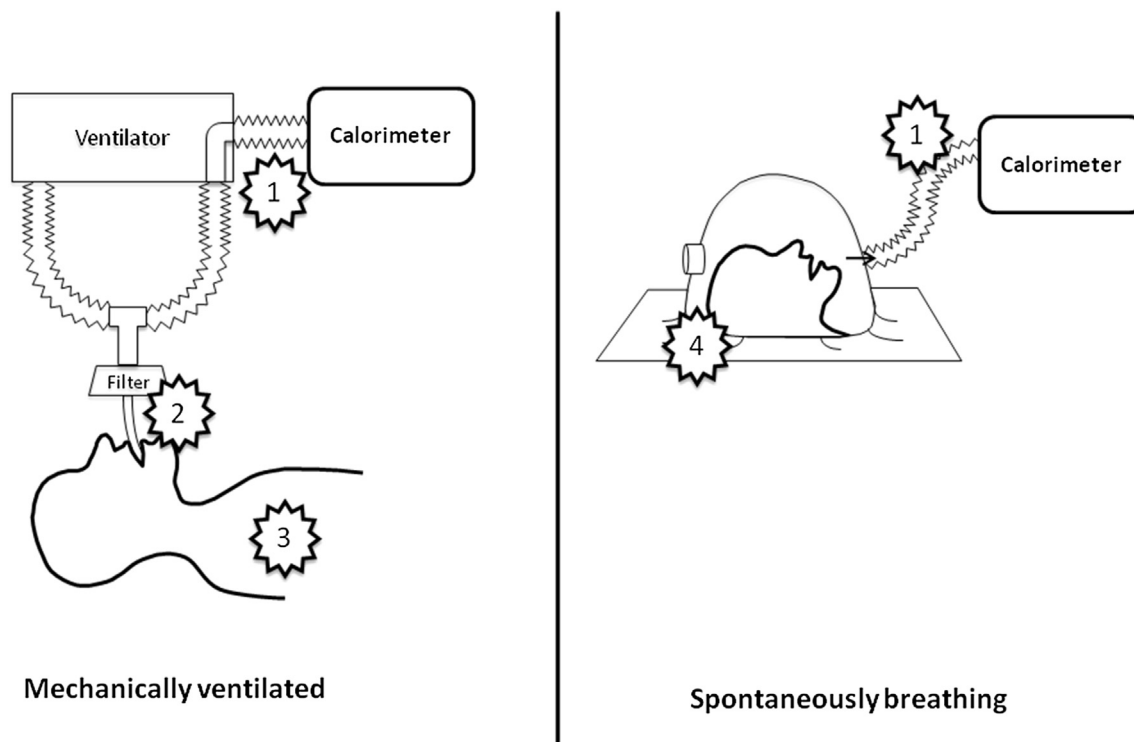


Fig. 2. Sources of air leaks during indirect calorimetry in spontaneously breathing patients and on those on mechanical ventilation. The avoidance of respiratory gas leaks is crucial to the accuracy of the energy expenditure measurement. 1: Tube connections with gas collection devices and calorimeters must be “air-tight”. 2: For patients on mechanical ventilation, leaks from the cuff of the endotracheal tube must be detected, as they can be significant in cases of high airway pressure. 3: Pathologies (e.g. bronchial fistula) and treatments (e.g. chest drain) causing air leaks from the lung must be detected. 4: Canopy and drape must be inspected for cracks and tears, and fitting tightly to each other. The drape should fully cover the surroundings of the canopy to avoid leaks.

Table 6

The Fick method (thermodilution) and related equations.

Calculation of O₂ content in blood

$$CaO_2 = (Hb) \times 1.38^{\#} \times SaO_2 + (0.003 \times PaO_2)$$

$$CvO_2 = (Hb) \times 1.38^{\#} \times SvO_2 + (0.003 \times PvO_2)$$

#: O₂ carrying capacity of Hb (1.34–1.39/gram, depending on literature)

Fick equation

$$VO_2 = (CaO_2 - CvO_2) \times CO \times 10 \text{ or}$$

$$VO_2 = 1.38 \times (Hb) \times (CO) \times (SaO_2 - SvO_2) / 10$$

Ca(v)O₂: content of O₂ in arterial (venous) blood, Sa(v)O₂: O₂ saturation of arterial (venous) blood, Pa(v)O₂: partial pressure of O₂ in arterial (venous) blood, CO: cardiac output (L/min).

3.2. Practical recommendations of clinical use

IC is successful when an appropriate device is used in optimal conditions, and the results are analyzed by experienced professionals in order to individualize the nutrition care. Although,

Table 7

Clinical situations requiring careful interpretation of energy expenditure measured by indirect calorimetry [26,27].

- Physical agitation or unstable sedation and/or analgesia
- Air leaks (>10% of minute volume)
- Unstable body temperature (>±1 °C change over last 1 h)
- Unstable pH (>±0.1 change over last 1 h)
- Oxygen enrichment (FiO₂ >60%)
- Organ support therapies: renal replacement or liver support therapy (pH alterations when conducted intermittently), ECMO (direct O₂ supply to the blood and CO₂ removal from the blood)

FiO₂: fraction of inspired oxygen, ECMO: extracorporeal membrane oxygenation.

these conditions are not easily met, practical recommendations are proposed below according to patient characteristics. Table 8 summarizes the important checkpoints for a successful routine use of IC. It can easily be adapted to create protocols after adjustment for the local medical practices.

3.3. Calorimetry: important considerations

IC is the only easy-to-use, non-invasive method to measure the EE of healthy active or inactive subjects, or of patients with various levels of metabolic stress in order to obtain immediate results [3,38]. Nevertheless, the lack of sufficient knowledge to interpret the results generated by IC may lead to erroneous prescription. The conditions of IC measurement are of paramount importance. The general statement is: the more stable the clinical situation, the more reliable the IC results. Whenever a situation is changing, IC should be repeated. For instance, IC obtained during the early phase of a critical illness should be repeated within the next 24–48 h to obtain a result reflecting the dynamic evolution of the disease [Fig. 3].

4. Developments of indirect calorimetry

4.1. The global initiative to promote calorimetry

Commercially available calorimeters are usually of large size and heavy weight, need time-consuming warm-up and calibration before measurement, require PCs to record and analyze results, require cumbersome disinfection of the device and repeated-use components after measurements, and are sold at relatively high costs [27,28]. The best way to promote IC is to make

available an easy-to-use, accurate and affordable device for daily use in in- and out-patients. The requirements of an ideal calorimeter have been defined using a bottom-up process [Table 9]. As current calorimeters were unable to meet the requirements defined by physicians, the International Multicentric Study Group for Indirect Calorimetry (ICALIC) was formed to develop and test an optimal device with the financial support of two international academic societies (The European Society of Intensive Care Medicine (ESICM) and The European Society for Clinical Nutrition and Metabolism (ESPEN)).

The new calorimeter has been validated against the gold standard technology for gas composition measurements, i.e. mass spectrometer (MAX300-LG, Extrel, Pittsburgh, USA). The mass spectrometer was specially tuned for breath gas analysis, to be able to measure O₂ and CO₂ with resolution up to 10 ppm and accuracy of $\pm 0.0025\%$. The analyzers for O₂ and CO₂ were evaluated for their accuracy in static concentration measurements using precision gas mixtures to simulate various clinically relevant O₂ (16%–21% for canopy measurements, and up to 70% for ventilator measurements) and CO₂ (0.3–5.0%) concentrations. Accuracy and response to dynamic concentration changes were evaluated in in-vivo evaluation in volunteers, by direct comparison of measured O₂ and CO₂ concentrations. The overall performance of the calorimeter consisting of the newly developed dynamic mixing chamber was tested by the

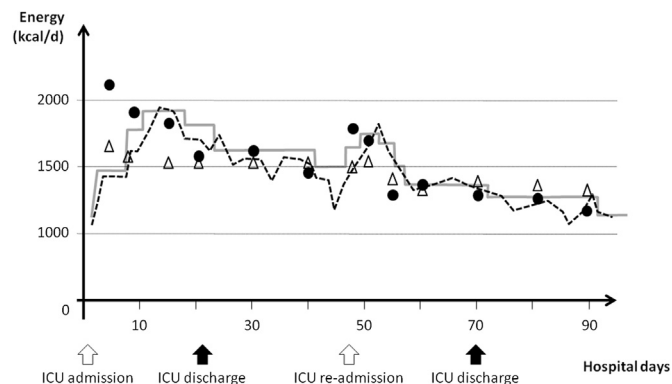


Fig. 3. Evolution of energy expenditure in critically ill patients. The evolution of energy expenditure (EE) of critically ill patients is dynamic according to the phase and the severity of the illness, treatment and extended bed rest. An illustration of such evolution in a septic patient with relapsed of the disease is presented. Dynamic change of EE during the early phase, and the difference in the evolution of EE during the first onset and the relapse due to factors such as bed rest and immobilization in the late phases are impossible to estimate accurately by predictive equations [11]. (Circle: EE by indirect calorimetry (kcal/d); triangle: EE by Faisy's equation; grey line (kcal/d): prescribed energy (kcal/d); dotted line: delivered energy).

Table 8

Checkpoints for successful indirect calorimetry.

	Mechanically ventilated	Spontaneously breathing
<i>Planning measurement</i>		
1. Frequency	Conduct calorimetry within 3–4 days after admission Repeat calorimetry every 2–3 days during the ICU stay Repeat calorimetry in case of changes in patient or disease conditions	
<i>Unsuitable conditions</i>		
2. Respiration	FiO ₂ >60% PEEP >10 cm H ₂ O Peak airway pressure >30 cm H ₂ O	O ₂ enrichment: Difficult with canopy Possible with leak-tight O ₂ mask using breath by breath device Intolerance for canopy and/or facemask
3. Agitation	Unstable sedation and/or analgesia Uncontrolled seizure and/or involuntary movements	
4. Treatments	Air leaks from ventilator circuit and/or endotracheal tube cuff Air leaks from chest drains Special consideration: Renal replacement therapy, liver support therapy, ECMO	
5. Immediate changes (<60 min before IC)	> ± 1 °C change of body temperature Change of drug dose: catecholamine, sedatives, analgesics, etc. Invasive procedures, mobilization, physical exercise	
<i>Before measurement</i>		
6. Device	Warm up and calibration (as required) Secure connections of tubes and components Search for any air leaks	
7. Feeding status	Continuous feeding preferred If fed, record: energy prescription and intake, duration (hrs) since last meal	Fasting preferred for out-patients (>8 h before IC)
8. Environment	Record: ventilation setting Maintain room temperature at 20–25 °C Ensure comfortable body position	Adjust canopy ventilation to maintain FeCO ₂ 0.8–1.2%
<i>During and after measurement</i>		
9. Quality of Measurement	Duration: 30 min or until stable state (calculated CV ^a <5% for VO ₂ and VCO ₂ for >5 min, CV of <10% for 25 min) RQ: <0.7 and >1.0 may suggest inadequate measurement Record: - agitation and body movements - any events affecting breathing pattern - changes in vasoactive drugs	
10. Disinfection	Disinfect device and components in contact with patients Discard single use components	

^a Coefficient of variation.

conventional mixing chamber method, using the mass spectrometer as the gas analyzer. The practical characteristics will be evaluated in a multicenter study which will start during the 1st semester of 2016, to ensure that the device is easy to use and fits the conditions found in various clinical settings.

Market release of the device is anticipated for 2017. Training courses are organized by ESPEN as part of the Life Long Learning (LLL) Courses and will be multiplied to allow optimal use of IC.

5. Why you should use indirect calorimetry

5.1. Rationale for measuring energy expenditure by indirect calorimetry

Energy expenditure of a patient is massively influenced by a number of intrinsic and extrinsic factors [Table 10] [19]. These factors have synergic or antagonist impact on the EE level and the estimation of the EE using a predictive equation based on anthropometric characteristics (i.e. body weight, height, gender, and age) is frequently inaccurate [3,20,39]. The use of multiplicative factor usually called “stress factor” has been proven to further deteriorate the estimation of EE based on equations. For example, obese patients present significant EE variations due to their underlying illnesses, variable body composition and degree of malnutrition [40]. Patients with chronic obstructive pulmonary disease or those with cancer have an elevated EE, which can be easily underestimated by predictive formula [41–43]. Critically ill patients with trauma or sepsis have dynamic changes of their EE during the successive phases of their critical illness [3,44–47]. Although much effort has been made to create predictive equations adapted to the clinical evolution of acute illness [48], IC remains the gold standard to measure EE [3,19]. The full benefits of nutrition support may be expected only if the patient specific EE is reflected in the nutrition prescription, according to the changes that occur during the course of the illness.

Table 9
Characteristics of the new indirect calorimeter defined by a bottom-up process of development.

Characteristics	Description
<i>Accuracy</i>	
Gas analyzers	±0.02% for O ₂ and CO ₂ (after calibration)
Flow analyzer	±2% (after calibration)
<i>Ease-of-use</i>	
Portable	<2 kg, maximum foot print:15–30 cm
Interface	Intuitive software, user manual not required
Calibration	<ul style="list-style-type: none"> Gas analyzer: Automatic periodic calibration against room air (no calibration gas required) Flow analyzer: Automatic
<i>Measurement</i>	
Duration	<10 min for standard measurement
Recording	<ul style="list-style-type: none"> Local memory buffer Various exportation formats (Excel, CSV, etc)
Connectivity	Wireless or USB
Battery operated	Up to 10 measurements (duration 20 min), 4hrs (continuous measurement)
<i>Safety</i>	
Approval	EC certification
Disinfection	<ul style="list-style-type: none"> Device covered by easy to clean material Single use components for patient contact (sampling tube, flow meter)
Compatible	Hospital devices
<i>Availability</i>	
Cost	<10'000 US \$
Market	Worldwide

5.2. Is measured EE always reflecting the energy needs?

This critical question has been asked many times and is frequently investigated, but the answer remains controversial. In general, the measured EE defines the energy target for the prescription of nutrition. However, during the early phase of an acute illness, endogenous energy supply covers most of the energy needs, a condition that is marginally affected by exogenous energy supplementation [Fig. 4] [49,50]. The energy administered may then massively exceed the requirements and generate relative overfeeding [3], a condition associated with deleterious consequences [Table 11] and poor outcome. This transitory period generally ends as soon the patient's overall condition improves. However, the value of IC measurements to evaluate the evolution of endogenous energy production needs further investigation. Therefore, careful interpretation of EE by IC in this phase is necessary for the adequate prescription of energy to avoid overfeeding. However, excessive restriction of energy will result in underfeeding, which has been associated with progressive loss of lean body mass [51], leading to poor outcomes. It should also be noted that predictive equations will not be able to take into account this type of metabolic alteration, and the degree of error in the estimation of EE will be unpredictable.

5.3. Respiratory quotient: another advantage of indirect calorimetry

IC allows for non-invasive measurement of EE in spontaneously breathing patients or those on mechanical ventilation, with or without O₂ enrichment [9,22,25,28]. An advantage of IC over other methods to measure EE is the capacity to derive the respiratory quotient (RQ) from direct measurements. The RQ corresponds to the quotient of VCO₂ and VO₂ ($RQ = VCO_2/VO_2$) [25,28], which enables the calculations of the substrate oxidation rates for glucose and lipids. This would especially allow detecting net lipogenesis. For patients with chronic illnesses, EE reflects the energy needs while the RQ reflects the composition of oxidized substrates [5]. This information is helpful to tailor the prescription of the nutrition regimen [28] by observing the match between the energy intake and the food quotient, i.e. the RQ of the energy substrates according to their food composition.

For critically ill patients, it allows to visualize the metabolic alterations, especially during the early phase. IC measurements should be repeated to monitor the dynamic changes, and to optimize the prescription of energy [3].

5.4. Routine use of indirect calorimetry

IC is rarely routinely used in medical institutions across the world [7,25] in spite of its value for a wide range of patients. Such a limited use of IC is mainly due to the unavailability of calorimeters, the insufficient awareness about the impact of optimal nutrition support on the patients outcome [20], the lack of expertise for interpretation of results, costs of device and related manpower. This section aims at clarifying these issues.

5.5. Critical illness

Patients in the ICU for >4 days or those after major surgery are good candidates for IC as they undergo severe stress related to variable metabolic needs [52]. Indeed, these patients are at high nutritional risk, as they are unable to resume sufficient oral intake instantly and often require enteral or parenteral nutrition [7,53,54]. Studies in critically ill patients have repeatedly reported gross underfeeding during the ICU stay [55]. Various factors such as gut

Table 10
Factors influencing energy expenditure.

- Age, sex, body height, body mass, body temperature Brain activity, endocrine profile, systemic inflammation
- Muscle contractions or paralysis, physical activity
- Fasting or post-absorptive state
- Environmental temperature
- Drugs (e.g. alpha adrenergic stimulant, beta-blockers, sedatives, muscle relaxants)

intolerance and treatment interventions delay full enteral feeding, resulting in insufficient energy provision. Underfeeding is closely associated with higher complication rates and poor outcomes [13,17,56–59]. Overfeeding has also been repeatedly associated with poor outcome and results often from the use of predictive equations [60]. Recent evidence points out the importance of optimal nutrition starting within 3–4 days after ICU admission [16,18,20,61–63], at a time when predictive equations are exceedingly unreliable due to the variable responses of individual patients to the critical illness [8]. In other words, optimal nutrition promotes better clinical outcome and IC is necessary to tailor the prescription to the real needs of the patient [62–64].

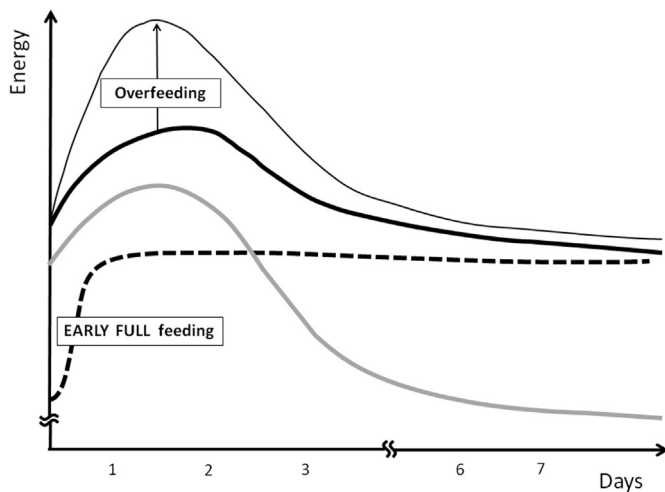


Fig. 4. Conceptual presentation of the relative overfeeding frequently related to parenteral nutrition during the early phase of critical illness. During the acute phase of the critical illness, the release of endogenous energy substrates is increased and meets total energy expenditure (TEE), and administering energy does not immediately terminate this response. Introducing full feeding in this early phase usually results in overfeeding, as the endogenous energy production is not attenuated by energy administration thus creates an excessive energy source above TEE. (Solid bold line: Total energy expenditure; grey bold line: adapted endogenous energy production; dotted bold line: early energy administration; thin line: combined endogenous and exogenous energy administration).

Table 11
Effects of overfeeding and underfeeding.

	Insufficient energy intake	Excessive energy intake
Early signs	Hypoglycemia Hypothermia	Hyperglycemia Hyperlipidemia (triglycerides) Hypercapnea
Delayed signs	Infectious complications Impaired immunity Impaired healing Loss of lean and fat body mass Impaired muscle function	Infectious complications Impaired immunity Liver steatosis Increased fat mass

The course of EE of severely ill patients features dynamic changes as a consequence of stress, prolonged bed rest, atrophy of the metabolically active lean tissue mass (i.e. 300–600 g of tissue/day), medications (catecholamine, sedatives, neuromuscular blocking agents, etc) [44,65–68], and modified by mechanical organ support therapies such as mechanical ventilation, renal replacement and liver support therapies. Thus, IC should be repeated as the clinical condition changes to accurately define the energy target [16,63].

The obese patients constitute an increasing proportion of the ICU patient population. Their energy requirements are particularly poorly addressed by predictive equations [19]. IC is the only way to determine their metabolic requirements accurately.

In summary, it is recommended to perform IC on days 3 or 4 after ICU admission, major surgery or trauma in order to set the energy target [Fig. 5].

5.6. In patients and outpatients with chronic conditions

Patients with chronic conditions are good candidates for IC, although their changes of EE are not as dynamic as in ICU or surgical patients. Indeed, chronic diseases or treatments modify the metabolically active lean body mass and the level of daily physical activity, which in turn significantly alter the energy needs and challenge the estimation of EE by predictive equations. Typically, important modifications of the body composition or of the physical activity deeply influence EE. Table 12 shows the most common pathologies with important EE alterations.

IC is necessary to confirm the energy expenditure and optimize the recommendation for food intake or the prescription of nutrition support. Repetition of IC should be considered according to the appearance of substantial modification of the patient status. Conducting IC together with the measurement of the body weight and the body composition is useful to further optimize the nutrition prescription by observing the effect of energy intake on these parameters [25,74].

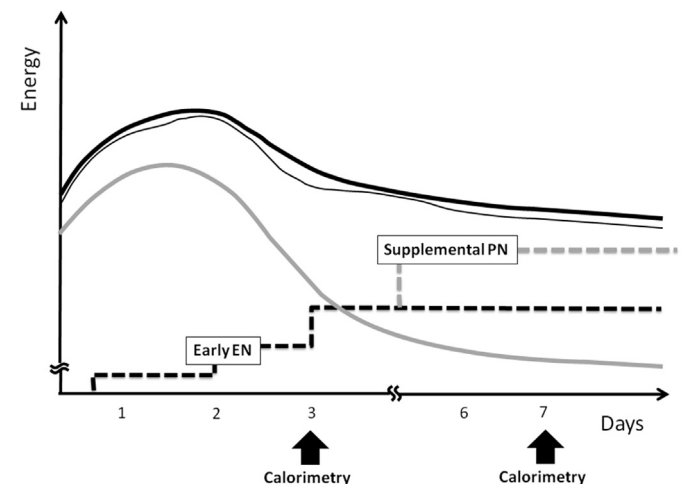


Fig. 5. Conceptual presentation of optimal feeding strategy to avoid both overfeeding and underfeeding in critical illness: Introducing the adequate amount of feeding in proportion to the body's capacity to down-regulate endogenous substrate production avoids both early overfeeding and late underfeeding. Repeated calorimetry is needed to monitor the dynamic changes of energy expenditure, however, providing the optimal amount of energy still requires special attention to avoid both underfeeding and overfeeding. (Solid bold line: Total energy expenditure; grey bold line: adapted endogenous energy production; dotted bold line: energy administration by EN; grey dotted bold line: energy administration by PN; thin line: combined endogenous and exogenous energy administration).

Table 12
Common chronic pathologies and treatments with important alterations of energy expenditure.

Conditions	Effects on energy expenditure
<i>Respiratory diseases</i>	
COPD	↑ Increased respiratory efforts [69]
Cystic fibrosis	↑ [70]
<i>Metabolic diseases</i>	
Adrenal gland disease	↑ or ↓ Increased release of catecholamine [71]
Thyroid diseases	↑ or ↓ Unpredictable change after surgical treatment
<i>Muscle tone alteration</i>	
Neuromuscular degenerative diseases	↓ Degeneration and disuse of muscle tissue
Paralysis	↓ Disuse and atrophy of paralyzed body area
Seizure, involuntary movements	↑ Increased muscle activity [73]
<i>Cachexic conditions</i>	
Cancer	↑ or ↓ Cancer growth and inflammation
AIDS	↑ or ↓ Progressive reduction of lean body mass
Cardiomyopathy	↓ Chronic infection and inflammation
<i>Malnutrition</i>	
Obesity	↑ or ↓ Progressive cachexia
Anorexia	↓ Progressive reduction of lean body mass
<i>Organ support therapies</i>	
Hemodialysis/peritoneal dialysis	↑ or ↓ Chronic inflammation
Continuous positive airway pressure (CPAP)	↑ or ↓ Progressive reduction of lean body mass
	Increased respiratory efforts, modified by mechanical support

5.7. Impact of IC on patient care and hospital economy

Malnutrition is associated with increased morbidity, length of stay and costs [75]. Oral nutritive supplements, and enteral and parenteral nutrition are related with improved outcome, but both underfeeding and overfeeding have been shown to mitigate the impact of nutrition support [63,76,77]. The prescription of nutrition therapy aims at matching the energy target as defined by predictive formulas. Unfortunately, these formulas are often inaccurate. Therefore, we hypothesize that promoting a large-scale use of IC to measure EE of in- and outpatients should optimize nutrition care, clinical outcome and costs.

6. Conclusion

Calorimetry is needed to optimize nutrition care for patients with various clinical conditions. The use of calorimetry is currently limited by various setbacks, mainly related to the lack of an adequate device. An ongoing initiative to develop a new calorimeter is expected to provide practical solutions for the current limitations, and make available a calorimeter corresponding to the requirements by clinicians for in- and outpatients, featuring accuracy, ease-of-use and affordable cost. Online and live educational courses will further mount the optimal use of calorimetry.

Conflict of interest statement

All authors have declared that they have no conflict of interest related to this project.

Statement of authorship

Taku Oshima and Claude Pichard have outlined this manuscript, which was developed, enriched, reviewed and approved by each of the co-authors.

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References

- [1] Bursztein SED, Askanazi JA, Kinney JM. Energy metabolism, indirect calorimetry, and nutrition. Baltimore, Maryland, USA: Williams and Wilkins; 1989.
- [2] Weir JB. New methods for calculating metabolic rate with special reference to protein metabolism. *J Physiol* 1949;109:1–9.
- [3] Fraipont V, Preiser JC. Energy estimation and measurement in critically ill patients. *J Parenter Enter Nutr* 2013;37:705–13.
- [4] Wilmore JH, Costill DL. Adequacy of the Haldane transformation in the computation of exercise VO₂ in man. *J Appl Physiol* 1973;35:85–9.
- [5] Frayn KN. Calculation of substrate oxidation rates in vivo from gaseous exchange. *J Appl Physiol Respir Environ Exerc Physiol* 1983;55:628–34.
- [6] Kyle UG, Arriaza A, Esposito M, Coss-Bu JA. Is indirect calorimetry a necessity or a luxury in the pediatric intensive care unit? *J Parenter Enter Nutr* 2012;36:177–82.
- [7] De Waele E, Spapen H, Honore PM, Mattens S, Van Gorp V, Diltor M, et al. Introducing a new generation indirect calorimeter for estimating energy requirements in adult intensive care unit patients: feasibility, practical considerations, and comparison with a mathematical equation. *J Crit Care* 2013;28(884):e881–886.
- [8] McClave SA, Lowen CC, Kleber MJ, Nicholson JF, Jimmerson SC, McConnell JW, et al. Are patients fed appropriately according to their caloric requirements? *J Parenter Enter Nutr* 1998;22:375–81.
- [9] Graf S, Karsegard VL, Viatte V, Heidegger CP, Fleury Y, Pichard C, et al. Evaluation of three indirect calorimetry devices in mechanically ventilated patients: which device compares best with the Deltatrac II((R))? A prospective observational study. *Clin Nutr* 2015;34:60–5.
- [10] Preiser JC, van Zanten AR, Berger MM, Biolo G, Casaer MP, Doig GS, et al. Metabolic and nutritional support of critically ill patients: consensus and controversies. *Crit Care* 2015;19:35.
- [11] Faisy C, Guerot E, Diehl JL, Labrousse J, Fagon JY. Assessment of resting energy expenditure in mechanically ventilated patients. *Am J Clin Nutr* 2003;78:241–9.
- [12] Wang Z, Heshka S, Gallagher D, Boozer CN, Kotler DP, Heymsfield SB. Resting energy expenditure-fat-free mass relationship: new insights provided by body composition modeling. *Am J Physiol Endocrinol Metab* 2000;279:E539–45.

- [13] Faisy C, Candela Llerena M, Savalle M, Mainardi JL, Fagon JY. Early ICU energy deficit is a risk factor for Staphylococcus aureus ventilator-associated pneumonia. *Chest* 2011;140:1254–60.
- [14] Ekpe K, Novara A, Mainardi JL, Fagon JY, Faisy C. Methicillin-resistant Staphylococcus aureus bloodstream infections are associated with a higher energy deficit than other ICU-acquired bacteremia. *Intensive Care Med* 2014;40:1878–87.
- [15] Casar MP, Mesotten D, Hermans G, Wouters PJ, Schetz M, Meyfroidt G, et al. Early versus late parenteral nutrition in critically ill adults. *N Engl J Med* 2011;365:506–17.
- [16] Singer P, Anbar R, Cohen J, Shapiro H, Shalita-Chesner M, Lev S, et al. The tight calorie control study (TICACOS): a prospective, randomized, controlled pilot study of nutritional support in critically ill patients. *Intensive Care Med* 2011;37:601–9.
- [17] Petros S, Horbach M, Seidel F, Weidhase L. Hypocaloric vs normocaloric nutrition in critically ill patients: a prospective randomized pilot trial. *J Parenter Enter Nutr* 2014;40:242–9.
- [18] Wei X, Day AG, Ouellette-Kuntz H, Heyland DK. The association between nutritional adequacy and long-term outcomes in critically ill patients requiring prolonged mechanical ventilation: a multicenter cohort study. *Crit Care Med* 2015;43:1569–79.
- [19] Frankenfield DC, Ashcraft CM. Estimating energy needs in nutrition support patients. *J Parenter Enter Nutr* 2011;35:563–70.
- [20] Guttormsen AB, Pichard C. Determining energy requirements in the ICU. *Curr Opin Clin Nutr Metab Care* 2014;17:171–6.
- [21] Sundstrom M, Tjader I, Rooyackers O, Wernerman J. Indirect calorimetry in mechanically ventilated patients. A systematic comparison of three instruments. *Clin Nutr* 2013;32:118–21.
- [22] Graf S, Karsgaard VL, Viatte V, Maisonneuve N, Pichard C, Genton L. Comparison of three indirect calorimetry devices and three methods of gas collection: a prospective observational study. *Clin Nutr* 2013;32:1067–72.
- [23] Black C, Grocott MP, Singer M. Metabolic monitoring in the intensive care unit: a comparison of the Medgraphics Ultima, Deltatrac II, and Douglas bag collection methods. *Br J Anaesth* 2014;114:261–8.
- [24] Sundström M, Fiskaare E, Tjäder I, Norberg Å, Rooyackers O, Wernerman J. Measuring Energy Expenditure in the Intensive Care Unit: a comparison of indirect calorimetry by E-sCOVX and Quark RMR with Deltatrac II in mechanically ventilated critically ill patients. *Crit Care* 2016. <http://dx.doi.org/10.1186/s13054-016-1232-6>.
- [25] Psota T, Chen KY. Measuring energy expenditure in clinical populations: rewards and challenges. *Eur J Clin Nutr* 2013;67:436–42.
- [26] Compher C, Frankenfield D, Keim N, Roth-Yousey L. Best practice methods to apply to measurement of resting metabolic rate in adults: a systematic review. *J Am Diet Assoc* 2006;106:881–903.
- [27] AARC. AARC Clinical Practice Guideline. Metabolic measurement using indirect calorimetry during mechanical ventilation. 2004 Revision & Update. *Respir Care* 2004;49:1073–9.
- [28] Haugen HA, Chan LN, Li F. Indirect calorimetry: a practical guide for clinicians. *Nutr Clin Pract* 2007;22:377–88.
- [29] Mansell PI, Macdonald IA. Reappraisal of the Weir equation for calculation of metabolic rate. *Am J Physiol* 1990;258:R1347–54.
- [30] Takala J, Keinanen O, Vaisanen P, Kari A. Measurement of gas exchange in intensive care: laboratory and clinical validation of a new device. *Crit Care Med* 1989;17:1041–7.
- [31] Zhao D, Xian X, Terrera M, Krishnan R, Miller D, Bridgeman D, et al. A pocket-sized metabolic analyzer for assessment of resting energy expenditure. *Clin Nutr* 2014;33:341–7.
- [32] Stapel SN, de Grooth HJ, Alimohamad H, Elbers PW, Girbes AR, Weijs PJ, et al. Ventilator-derived carbon dioxide production to assess energy expenditure in critically ill patients: proof of concept. *Crit Care* 2015;19:370.
- [33] Sandstrom R, Drott C, Hyltander A, Arfvidsson B, Schersten T, Wickstrom I, et al. The effect of postoperative intravenous feeding (TPN) on outcome following major surgery evaluated in a randomized study. *Ann Surg* 1993;217:185–95.
- [34] Phang PT, Rich T, Ronco J. A validation and comparison study of two metabolic monitors. *J Parenter Enter Nutr* 1990;14:259–61.
- [35] Tissot S, Delafosse B, Bertrand O, Bouffard Y, Viale JP, Annat G. Clinical validation of the Deltatrac monitoring system in mechanically ventilated patients. *Intensive Care Med* 1995;21:149–53.
- [36] El-Orbany M, Salem MR. Endotracheal tube cuff leaks: causes, consequences, and management. *Anesth Analg* 2011;117:428–34.
- [37] De Waele E, van Zwam K, Mattens S, Staessens K, Diltsoer M, Honore PM, et al. Measuring resting energy expenditure during extracorporeal membrane oxygenation: preliminary clinical experience with a proposed theoretical model. *Acta Anaesthesiol Scand* 2015;59:1296–302.
- [38] De Waele E, Spapen H, Honore PM, Mattens S, Rose T, Huyghens L. Bedside calculation of energy expenditure does not guarantee adequate caloric prescription in long-term mechanically ventilated critically ill patients: a quality control study. *ScientificWorldJournal* 2012;2012:909564.
- [39] Frankenfield DC, Coleman A, Alam S, Cooney RN. Analysis of estimation methods for resting metabolic rate in critically ill adults. *J Parenter Enter Nutr* 2009;33:27–36.
- [40] Mogensen KM, Andrew BY, Corona JC, Robinson MK. Validation of the society of critical care medicine and American society for parenteral and enteral nutrition recommendations for caloric provision to critically ill obese patients: a pilot study. *J Parenter Enter Nutr* 2015;40:713–21.
- [41] Ramires BR, de Oliveira EP, Pimentel GD, McLellan KC, Nakato DM, Faganello MM, et al. Resting energy expenditure and carbohydrate oxidation are higher in elderly patients with COPD: a case control study. *Nutr J* 2011;11:37.
- [42] Lieffers JR, Mourtzakis M, Hall KD, McCargar LJ, Prado CM, Baracos VE. A viscerally driven cachexia syndrome in patients with advanced colorectal cancer: contributions of organ and tumor mass to whole-body energy demands. *Am J Clin Nutr* 2009;89:1173–9.
- [43] Fearon K, Strasser F, Anker SD, Bosaeus I, Bruera E, Fainsinger RL, et al. Definition and classification of cancer cachexia: an international consensus. *Lancet Oncol* 2011;12:489–95.
- [44] Puthucherry ZA, Rawal J, McPhail M, Connolly B, Ratnayake G, Chan P, et al. Acute skeletal muscle wasting in critical illness. *Jama* 2013;310:1591–600.
- [45] Hartl WH, Jauch KW. Metabolic self-destruction in critically ill patients: origins, mechanisms and therapeutic principles. *Nutrition* 2014;30:261–7.
- [46] Cerra FB, Siegel JH, Coleman B, Border JR, McMenemy RR. Septic autotocannibalism. A failure of exogenous nutritional support. *Ann Surg* 1980;192:570–80.
- [47] Preiser JC, Ichai C, Orban JC, Groeneveld AB. Metabolic response to the stress of critical illness. *Br J Anaesth* 2014;113:945–54.
- [48] Savard JF, Faisy C, Lerolle N, Guerot E, Diehl JL, Fagon JY. Validation of a predictive method for an accurate assessment of resting energy expenditure in medical mechanically ventilated patients. *Crit Care Med* 2008;36:1175–83.
- [49] Wolfe RR. Sepsis as a modulator of adaptation to low and high carbohydrate and low and high fat intakes. *Eur J Clin Nutr* 1999;53(Suppl. 1):S136–42.
- [50] Rennie MJ. Anabolic resistance in critically ill patients. *Crit Care Med* 2009;37:S398–9.
- [51] Hoshino E, Pichard C, Greenwood CE, Kuo GC, Cameron RG, Kurian R, et al. Body composition and metabolic rate in rat during a continuous infusion of cachectin. *Am J Physiol* 1991;260:E27–36.
- [52] Finnerty AM, Mabvuure NT, Ali A, Kozar RA, Herndon DN. The surgically induced stress response. *J Parenter Enter Nutr* 2013;37:215–9S.
- [53] Heyland DK, Dhaliwal R, Lemieux M, Wang M, Day AG. Implementing the PEP up protocol in critical care units in Canada: results of a multicenter, quality improvement study. *J Parenter Enter Nutr* 2014;39:698–706.
- [54] Hiesmayr M. Nutrition risk assessment in the ICU. *Curr Opin Clin Nutr Metab Care* 2012;15:174–80.
- [55] Alberda C, Gramlich L, Jones N, Jeejeebhoy K, Day AG, Dhaliwal R, et al. The relationship between nutritional intake and clinical outcomes in critically ill patients: results of an international multicenter observational study. *Intensive Care Med* 2009;35:1728–37.
- [56] Thibault R, Graf S, Clerc A, Delieuvin N, Heidegger CP, Pichard C. Diarrhoea in the ICU: respective contribution of feeding and antibiotics. *Crit Care* 2013;17:R153.
- [57] Heidegger CP, Romand JA, Treggiari MM, Pichard C. Is it now time to promote mixed enteral and parenteral nutrition for the critically ill patient? *Intensive Care Med* 2007;33:963–9.
- [58] Villet S, Chiolerio RL, Bollmann MD, Revelly JP, Cayeux RNM, Delarue J, et al. Negative impact of hypocaloric feeding and energy balance on clinical outcome in ICU patients. *Clin Nutr* 2005;24:502–9.
- [59] Dvir D, Cohen J, Singer P. Computerized energy balance and complications in critically ill patients: an observational study. *Clin Nutr* 2006;25:37–44.
- [60] Berger MM, Pichard C. Development and current use of parenteral nutrition in critical care - an opinion paper. *Crit Care* 2014;18:478.
- [61] Oshima T, Pichard C. Parenteral nutrition: never say never. *Crit Care* 2015;19(Suppl. 3):S5.
- [62] Weijs P, Looijaard W, Beishuizen A, Girbes A, Oudemans-van Straaten HM. Early high protein intake is associated with low mortality and energy over-feeding with high mortality in non-septic mechanically ventilated critically ill patients. *Crit Care* 2014;18:701.
- [63] Heidegger CP, Berger MM, Graf S, Zingg W, Darmon P, Costanza MC, et al. Optimisation of energy provision with supplemental parenteral nutrition in critically ill patients: a randomised controlled clinical trial. *Lancet* 2013;381:385–93.
- [64] Singer P, Hiesmayr M, Biolo G, Felbinger TW, Berger MM, Goeters C, et al. Pragmatic approach to nutrition in the ICU: expert opinion regarding which calorie protein target. *Clin Nutr* 2014;33:246–51.
- [65] Grosu HB, Lee YI, Lee J, Eden E, Eikermann M, Rose KM. Diaphragm muscle thinning in patients who are mechanically ventilated. *Chest* 2012;142:1455–60.
- [66] English KL, Paddon-Jones D. Protecting muscle mass and function in older adults during bed rest. *Curr Opin Clin Nutr Metab Care* 2010;13:34–9.
- [67] Honore PM, De Waele E, Jacobs R, Mattens S, Rose T, Joannes-Boyau O, et al. Nutritional and metabolic alterations during continuous renal replacement therapy. *Blood Purif* 2013;35:279–84.
- [68] Biolo G, Ciochi B, Lebenstedt M, Barazzoni R, Zanetti M, Platen P, et al. Short-term bed rest impairs amino acid-induced protein anabolism in humans. *J Physiol* 2004;558:381–8.
- [69] Rabinovich RA, Louvaris Z, Raste Y, Langer D, Van Remoortel H, Giavedoni S, et al. Validity of physical activity monitors during daily life in patients with COPD. *Eur Respir J* 2013;42:1205–15.

- [70] Frankenfield DC, Ashcraft CM, Drasher TL, Reid EK, Vender RL. Characteristics of resting metabolic rate in critically ill, mechanically ventilated adults with cystic fibrosis. *J Parenter Enter Nutr* 2015. <http://dx.doi.org/10.1177/0148607115617152>.
- [71] Xu WP, Cao DX, Lin ZM, Wu GH, Chen L, Zhang JP, et al. Analysis of energy utilization and body composition in kidney, bladder, and adrenal cancer patients. *Urol Oncol* 2012;30:711–8.
- [72] Spadafranca A, Cappelletti C, Leone A, Vignati L, Battezzati A, Bedogni G, et al. Relationship between thyroid hormones, resting energy expenditure and cardiometabolic risk factors in euthyroid subjects. *Clin Nutr* 2015;34:674–8.
- [73] Capecci M, Petrelli M, Emanuelli B, Millevolte M, Nicolai A, Provinciali L, et al. Rest energy expenditure in Parkinson's disease: role of disease progression and dopaminergic therapy. *Park Relat Disord* 2013;19:238–41.
- [74] Mourtzakis M, Wischmeyer P. Bedside ultrasound measurement of skeletal muscle. *Curr Opin Clin Nutr Metab Care* 2014;17:389–95.
- [75] Milte RK, Ratcliffe J, Miller MD, Crotty M. Economic evaluation for protein and energy supplementation in adults: opportunities to strengthen the evidence. *Eur J Clin Nutr* 2013;67:1243–50.
- [76] Schulman RC, Mechanick JL. Metabolic and nutrition support in the chronic critical illness syndrome. *Respir Care* 2012;57:958–77. discussion 977–958.
- [77] Hughes MJ, Harrison EM, Wigmore SJ. Energy expenditure after liver resection: validation of a mobile device for estimating resting energy expenditure and an investigation of energy expenditure change after liver resection. *J Parenter Enter Nutr* 2015. <http://dx.doi.org/10.1177/0148607115601969>.